OXYGEN ENHANCEMENT OF FREE RADICAL DAMAGE TO DNA. A COMPARISON OF RADIATION-INDUCED DEGRADATION WITH COPPER-CATALYZED DEGRADATION BY H₂O₂

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The best known concept to explain oxygen enhancement in irradiated aqueous solutions containing DNA and thiol is that of 'chemical repair' of ('OH-induced) DNA intermediates by thiols due to hydrogen donation, and competitive damage fixation in oxygenated solution due to peroxidation of DNA intermediates.¹⁻³ More recently it has been proposed that 'self-repair', involving electron transfer from disulphide radical anions (RSS⁻ R) to DNA intermediates may be of importance in irradiated oxygen-free DNA/thiol and DNA/disulphide systems;⁴ oxygen enhancement can be easily explained in this case as 'self-repair' inhibition, due to scavenging of RSS⁻ R and precursors (see Scheme A). Both modes of action of oxygen may possibly occur in irradiated systems. When DNA/thiol solutions are incubated with Cu^{2+} , H_2O_2 and ascorbate (at high concentration), a Fenton-type reaction of DNA-Cu(I) with H_2O_2 is thought to generate 'OH radicals immediately at the target.⁵⁶ The RSS⁻ R precursor RS' is scavenged by AH⁻ in this system,⁷ and oxygen enhancement is feasible only via the 'damage fixation' mode (Scheme B).

The results shown in Table 1 demonstrate a substantial oxygen effect in γ -irradiated DNA/GSH and DNA/GSSG solutions, but no oxygen enhancement is detectable in these systems when incubated with Cu²⁺, H₂O₂ and AH⁻, despite the higher GSH or GSSG concentration — (with gamma-irradiation a higher GSH level even promotes the oxygen effect).² It might be suggested that H₂O₂ does not generate OH radicals in reaction with DNA-Cu(I). An analysis of the kinetics and yield in system B

TABLE 1

Protective effect of glutathione (GSH and GSSG) in deoxygenated (N_2) and oxygenated (O_2) DNA solutions.

A) y-Radiolysis		$B) DNA-Cu(1) + H_2O_2$	
Additives (X)	PF	Additives (X)	PF
0,	0.95	 	0.89
$0.3 \mathrm{mM}\mathrm{GSH},\mathrm{N}_{2}$	12.8	1.5 mM GSH, N ₂	2.00
$0.3 \mathrm{mM}\mathrm{GSH},\mathrm{O}_{3}$	6.4	1.5 mM GSH. 0.	2.16
0.15 mM GSSG, N ₂	10.2	$1.5 \mathrm{mM}\mathrm{GSSG},\mathrm{N}_{2}$	1.81
$0.15 \mathrm{mM}\mathrm{GSSG},\mathrm{O}_2$	5.1	$1.5 \mathrm{mM}\mathrm{GSSG},\mathrm{O}_2$	2.22

A) 25 Gy/min ⁶⁰Co- γ -radiolysis of 0.1 g/1 DNA in 10 mM phosphate buffer; protection factors from dose(D)-effect curves: PF=D₁₇(X)/D₁₇(O), with D₁₇(O) = 250 Gy in oxygen-free solution.

B) Incubation at 22°C of 0.1 g/l DNA in 10 mM phosphate buffer with 10 μ M CuCl₂, 10 mM H₂O₂ and 20 mM AH⁻; protection factors from time (t) profiles of DNA degradation: PF=t₃₇(X)/t₃₇(O), with t₃₇(O) = 13.5 min in oxygen-free solution. After treatment A or B, 10 mM EDTA was added, and DNA degradation was tested fluorimetrically with ethidium bromide (see ref.⁶).

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A) Gamma-Radiolysis of DNA/RSH Solutions



B) Incubation of DNA/RSH with Cu^{2+} , H_2O_2 and Ascorbate





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CHEMICAL REPAIR

indicated however that OH is the damaging entity,⁶ and furthermore we have observed a characteristic fluorescence around 460 nm (under excitation at 350 nm) both in system A and B under hypoxic conditions, which can be assigned to OH-induced products. Therefore, to explain oxygen enhancement in γ -irradiated DNA/thiol(disulphide) solutions, the 'self-repair' concept apparently deserves further attention.

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